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Comparitive toxicity of cyclic peptides and depsipeptides in cultured rat hepatocytes

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Introduction: Low-molecular-weight (MW), cyclic popules (peptides linked through an amide linkage) and depsipeptides (peptides linked through an ester linkage) comprise a small group of metabolites produced by fungi, algae or bacteria. Among these cyclic peptides are cyclosporine (MW 1203), gramicidin-S (MW 1141) and microcystin-LR (MW 994), while valinomycis (MW 1111) and enniatin-B (MW 639) represent cyclic depsipeptides. These cyclic compounds possess varied pharmacological properties, ranging from antimicrobial activity FIP (valinousycia, enniatin-B, gramicidin-S, microcystin-LR) and strong immunosuppressive activity-[2] (cyclospurine), to antimalarial activity [3] (valinomycin, cyclosporine, gramicidin). Many of these small cyclic peptides possess ionophoric properties, exhibiting differences in ion selectivity and affinities [1]. -Mic "cat att

and affinities [1].

The toxicity (LD₂₀) of these compounds is in the range of microgram (microcystin-LR, 50 kg lip., mice) [2] to milligram (cyclosporine, 107 kg kg lip., mice) [2] quantities. Although mice treated with 200 ffg kg day of cyclosporine [3], or sublethal doses of microcystin-LR [3], developed hepatic vascular congestion and fatty liver, there is no information available on the hepatotoxicity of the other cyclic peptides and densipeptides. Microcystin-LR induces liver damage in mice [4] and necrosis of cultured hepatocytes after several hours of incubation with the toxin [4].

This study was designed to compare cell injury induced by these cyclic peptides and depsipeptides using the release of LDH and admine nucleotides from cultured hepatocytes,

Material and dischada: The following materials were obtained commercially from the indicated sources: gramicidin-S (Chemical Dynamics Corp., South Field, NJ), valinomycin (Calbiochem, La Jolin, CA), [MC]adenine (50 mCi mmol⁻¹, New England Nuclear, Boston, MA), tissue culture medium and foetal bevine serum albumin (GIBCO, Grand Island, NY), tissue culture were (Bectos-Dickinson Labware, Lincoln Park, NJ), rat tail collagen, collagenase type IV, 5'-adenosine monophosphate (AMP), 5'-adenosine diphosphate (ADP), 5'-adenosine triphosphate (ATP), 5'-inosine monophosphate (IMP), adenosine, and adenise (Sigma, St. Louis, MO).

Male FW.LEW, congenic, inbred rats (G. Anderson, USAMRIID, Fort Detrick, Frederick, MD) weighing between 250-300 g were used for all experiments. Microcystin-LR (> 95% purity by HPLC) was obtained from Dr W. Carnichael, Wright State University, Dayton, Ohio. The following materials were gifts from the indicated sources: cyclosporine (Sandoz Laburatories, East Hanover, NJ, and enniztin-B (> 95% purity by TLC) from Dr H.R. Burmeister, Northern Regional Research Center, USDA, Peoria, IL.

Rat hepatocytes were isolated and cultured according to

the method of Elliget and Kolaja [8]. After overnight incubation, hepatocytes were labelled with [14 C]adenine (0.2 µCi, 4 µM) as desc. bed by Shirhatti and Krishna [7]. Aliquots from the supernatants of prelabeled cells incubated with the cyclic peptides were analysed for the release of labelled-nucleotides and LDH at selected time intervals. The cells were lysed with 1 mL of 0.05% digitonin and an aliquat of the cell lysate was analysed for radioactivity (Beckman scintillation counter, model LS800, Fullerton, CA), LDH and protein content (Pierce protein reagent, Rockford, IL).

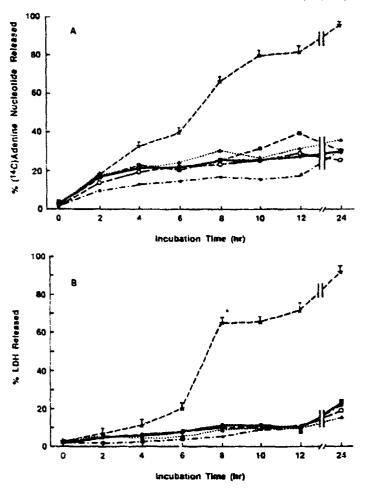
[14C]-Adenine 5 nucleotides (AMP, ADP, ATP, IMP), adenine and adenosine of hepatocyte supernatants were determined by thin layer chromatography on PEI-cellulose plates as described by Bockner and Ames [9] and compared with standards. The regions corresponding to the chromatographed standards were scraped from the plate and counted for radioactivity. Control and treated hepatocytes were examined under phase contrast microscope (Nikon Diaphot inverted phase contrast microscope) for morphological changes.

In conducting the research described in this report, the authors adhered to the Guide for Laboratory Animal Facilities and Core as promulgated by the Committee on the Guide for Laboratory Animal Resources, NAS/NRC.

Furthermore, microcystin-LR (0.1-50 µM) induced doseand time-dependent release of [14C]adenine nucleotides (Figure 2) and LDH (data not shown). In addition, I µM microcystin-LR caused deformation (rounding and blebbing) in cell morphology (data not shown) consistant with the observation of an earlier report [10].

In order to determine whether the other cyclic peptides and depsipeptides induced toxicity in cultured raf Repatocytes at levels > 10 µM, cells were incabated with 50 µM of valinomycin, cyclosporine, or grassicidin-S for a total of 24 h. Enniatin-B was not tested at 50 µM due to inadequate supplies. At 50 µM, valinomycin, cyclosporine and gramicidin-S induced a significant time-dependent release of both labelled-nucleotides and LDH from hepatocytes as compared to control cells (Figures 3A and 3B).

Differences in the percent of marker release were observed in cells treated with cyclic peptides and depaipeptide. Gramocidin-S, microcystin-LR or valinomycin treated cells released approximately 80% of the total nucleotides (Figure 3A) within the first 2 h of incubation. Cyclosporiae induced 90% release of labeled nucleotides from hepatocytes between 4-6 h of incubation (Figure 3A). The rate of LDH release



Figures IA and IB: Effect of I yM microcysin-LR (#—#), cyclosporine (0—0), gramicidin-S (#—#), valinomycin (8—#), or emiatin-B (A...A) on the release of [14 Cladenine nucleotides (IA) and LDH (IB) from cultured rat hepanocyses as compared to control (#—#). Each point represents the mean of six determinations, with 5-7% deviation. Values for the release of markers differed significantly from control figures only for microcystin-LR treatment after 4 h (p < 0.05, Student's t-test) Sandard deviation bars were eliminated for clarity.

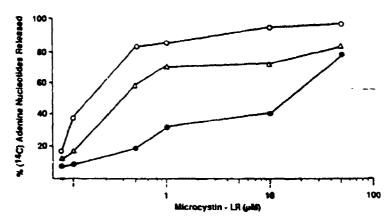
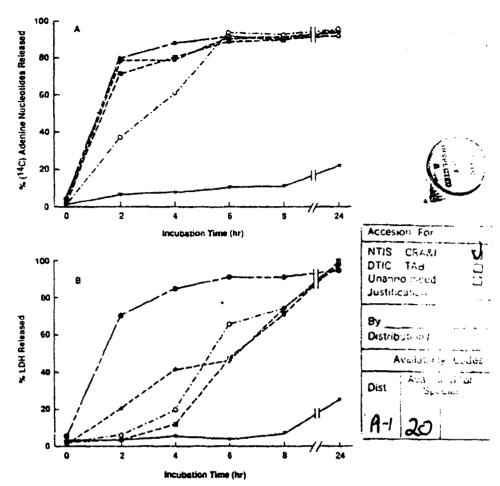


Figure 2: Effect of microcyzin-LR (0.1 – 50 µM) on the release of [*C]adenine inclassides from cultural rat hepetocytes incubated for 2 (0-0), 4 (0-0). Each point represents the mass of three determinations, with 2-3% deviation. Sandard deviation bars were eliminated for clarity.



Figures 3A and 3B: Effect of 50 μ M microcystin-LR (ψ — ψ), cyclosporine (ϕ — ϕ), gramicidin-S (ϕ — ϕ), or valinomycin (ϕ — ϕ) on the release of [**C]adenine nucleotides (3A) and LDH (3B) from cultured rat hepatocyses as compared to control (Ψ — Ψ). Each point represents the mean of six determinations, with λ -7% deviation. Values for the release of markers differed significantly (ϕ < 0.05, Student's (- test) from control figures for all treatments except for LDH release at 2 h in valinomycin and cyclosporme treat nens. Standard deviation bars were eliminated for clarity.

from gramicidin-S treated cells was parallel to the release of labelled nucleotides (Figures 3A and 3B).

There was a 2 h tag in the release of LDH as compared to the release of nucleotides from cells treated with validomycin or with cyclosporine. The percentages of LDI: released at 2 and 4 h from cells treated with microcystin-LR, valinomycin and cyclosporine were significantly less than the percentages of nucleotides released at the same time points (Figures 3A and 3B).

The Rf values for AMP, ADP, ATP, IMP and adenosine were 0.68, 0.34, 0.1, 0.58 and 0.54, respectively. Due to the poor resolution in separating adenosine from IMP, the bands corresponding to these two compounds were quantified as one and reported as IMP. The distribution of labelled nucleotides released into the medium (8 h, 50 µM) was the same in control and toxin treated cells (AMP, 89%; ADP, 8%; ATP, 0.5%).

In conclusion, the release of LDH and ademine nucleotides from cultured rat hopatocytes indicated that at 50 µM, the cyclic peptides and depsipeptides tested in this study were hepatotoxic. Comparatively, at low concentration (1 µM), microcystin-LR exhibited the greatest cytotoxicity among these hepatotoxins.

- i. Oschingikov, Y.A. and Ivanov, V.T. 1976. Mondalov Chem. J., 21. 18-25
- Thompson, a 15, 306-327 n, A.W., Whiting, P.H. and Simpson, J.G. 1984. Agents Actions,
- Bost, D., Desies, D. and Capton, A. 1983. Fed. Proc., 42, 1246 Runnegar, M.T. and Felconer, I.R. 1981. In: Carmichel, W.W., (ed.), The Water Environment: Algal Taxins and Health, pp. 325–342. Pleasan Press, New York
- Boland, J., Atkin, K., Britton, K., et al. 1984. Pushology, 16, 117-123. Found, T.L. and Samer Jr., J.J. 1981. In: Carmichael, W.W., (ed.), The me: Algel Toxins and Houth, pp. 365-387. Pleasan Press.
- esi, V. and Krishna, G. 1985. Analysical Binchess., 147, 410-418 Shirk
- Eligot, K.A. and Kolnin, G.J. 1963. J. Tieser Calture Methods, S. 1-6 Botherr, B.R. and Armes, B.N. 1962. J. Biol. Chem., 367, 9759-9769 Asse, T. and Jerg, K. 1986. J. Toxicol. Envir. 1981., 19, 325-336

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